

Difference Between Amplicor and Linnear Array Tests for Detecting HPV-Induced Cervical Abnormalities

Krivak Bolanča, Ines; Findri Guštek, Štefica; Oresčanin, Višnja

Source / Izvornik: **GJRA: Global Journal for Research Analysis, 2016, 5, 1 - 4**

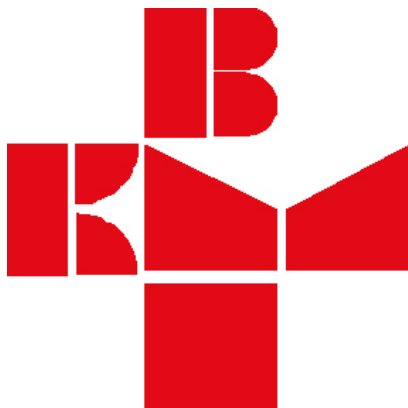
Journal article, Published version

Rad u časopisu, Objavljena verzija rada (izdavačev PDF)

Permanent link / Trajna poveznica: <https://um.nsk.hr/um:nbn:hr:264:182724>

Rights / Prava: [In copyright](#)/[Zaštićeno autorskim pravom.](#)

Download date / Datum preuzimanja: **2025-02-22**



Repository / Repozitorij:

[Merkur University Hospital Repository](#)

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/315786182>

Difference between Amplicor and Linnear Array tests for detecting HPV-induced cervical abnormalities

Article · March 2016

CITATIONS

0

READS

57

3 authors:



[Ines Krivak Bolanca](#)

Clinical Hospital Merkur

44 PUBLICATIONS 67 CITATIONS

[SEE PROFILE](#)

[Stefica Findri Gustek](#)

Ustanova za zdravstvenu skrb

33 PUBLICATIONS 368 CITATIONS

[SEE PROFILE](#)



[Visnja Orescanin](#)

ORESCANIN Ltd., A. Jaksica 30, Zagreb, Croatia, <http://orescanin.webnode.com/>

215 PUBLICATIONS 2,362 CITATIONS

[SEE PROFILE](#)



Difference Between Amplicor And Linnear Array Tests for Detecting Hpv- Induced Cervical Abnormalities

* Krivak-Bolanca

Unit for Gynaecologic Cytology ,Department of Clinical Cytology and Cytogenetics, Merkur University Hospital, Zagreb * Corresponding author

Findri Gustek S.

Outpatient Clinic for Gynecology, Urology and Occupational Medicine, dr. Findri Gustek, Zagreb, Croatia

Orescanin V.

ORESCANIN Ltd., A. Jaksica 30, Zagreb, Croatia

ABSTRACT

The aim of the study was to morphologically analyse cervical samples prepared with liquid-based cytology using transport medium residual samples collected for the detection of hr-HPV, and to genotype the samples by the reverse hybridization strip assay in each specimen. For detection of viral infection and detailed genotyping the Amplicor assay and Linnear Array methods were used. Cytological diagnoses were made according to the Bethesda classification. Cell abnormalities were observed in HPV positive samples, but also in samples that tested HPV negative. Detection of HPV infection showed infection with one viral type but in most cases infection was caused by more viral types. Between those two tests, difference was noticed, reflecting on the frequency and on the appearance of existing low and high grade cervical lesion. The sensitivity of these tests has been observed to be somewhat different, though not statistically significantly.

KEYWORDS : HPV, genotyping, cervical lesion, cytology

INTRODUCTION:

An infection with human papillomaviruses (HPV) is very common, especially among young women and adolescents. In spite of the generally benign outcome of these infections, certain types of HPV have the potential to transform cells and are associated with mucosal precanceroses or malignancies. The evidence of the importance of persistent human papillomavirus infection in women with cervical abnormalities has become very convincing^{1,2}.

The physicians' understanding of the natural history of HPV disease has significantly improved, due to the development of molecular methods and techniques. Detection of the infection relies on proving the viral DNA and for that purpose a range of different commercial and molecular tests have been developed³⁻⁵. The most used assay is HC2 (hybrid capture test) – a ready-to-use assay for routine diagnostics and detection of HPV infection, regardless of HPV specific types. Advantage of this method are: reproducibility, reliability and there is no need for specialized employees for performing diagnostic methods and presenting results. But, for clinical management of HPV-induced precancerous lesions is very important to be able to distinguish an individual type of HPV. For that, polymerase chain reaction (PCR) - based methods are used and shortly become a "gold standard" for detection of HPV infection. However, these methods are expensive; a completely equipped laboratory is required as well as educated employees for performing molecular methods and still different PCR methods shows different sensitivity and specificity. Amplicor HPV test (Roche Amplicor Human Papilloma Virus Test, Roche Diagnostic, Switzerland) - one of the standardized PCR-based tests for detection high risk HPV genotypes (hr-HPV) has been commercialised for detecting 16 different, hr-HPV genotypes. Currently, mostly used HPV genotyping tests are based on reverse hybridization of amplified HPV products on a membrane-bound probes, such is Linnear Array HPV Genotyping Test (Linneear Array Test, Roche Diagnostic, Switzerland).

The detection of the oncogenic, hr-HPV genotypes as well as an individual HPV genotyping had consequently become an important part of cervical carcinoma screening and detection of precancerous lesions. Aim of this study is to emphasize the importance of genotyping in women with the cytological abnormalities of the cervix and proven HPV infection.

EXAMINEES AND METHODS:

In this study, the cervical specimens from 339 patients were collected with the cervix brush (Cervex-Brush, Rovers Medical Devices) which was placed in the PreserveCyt vial (Cytyc Corporation, Hologic) and after making a monolayered smears with ThinPrep 2000 processor (Hologic), they were analyzed. In each of the patient HPV genotyping was done, the type of infection (mono- or multi-type infection) was determined, and the presence of particular stage of dysplasia was cytologically diagnosed using Bethesda classification.

In 269 patients a certain degree of cytological abnormalities were found, so supplementary smears were made from the vials of those patients. Cervical cytology specimens from 70 patients of the same age, collected at the same gynaecological out-patient clinic and during the same period without proven HPV infection and with normal cytological results, were used as a control group.

Residual solution in the PreserveCyt vials were used for detecting viral DNA by Amplicor Test as well as for individual genotyping with Linnear Array test.

Amplicor test was performed for detecting HPV infection in cervical cells collected in liquid media. It is a qualitative test that utilizes amplification of target DNA by the PCR and hybridization for the group detection of 13 hr HPV genotypes: 16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59 and 68, but does not allow identification of individual genotypes. The test uses biotinylated PGM1 primers to define a sequence of nucleotides within polymorphic L1 region of the HPV genome that is approximately 165 base pairs long designed to amplify HPV DNA from 13 hrHPV genotypes. Following PCR amplification and denaturation, detection occurs by hybridization of amplicons to probes coated in 96-microwells plates. Additionally, for detailed genotyping, the Linneear Array test was used. Linneear Array test is qualitative test, as well, and it utilizes amplification of target DNA by the PCR and hybridization for the individually detection of 37 anogenital HPV genotypes. The test uses biotinylated PGM1 primers to define a sequence of nucleotides within polymorphic L1 region of the HPV genome that is approximately 450 base pairs long and designed to amplify HPV DNA from 37 HPV genotypes. Following PCR amplification and denaturation, detection occurs by reverse hybridization of amplicons to immobilized membrane-bound probes.

All steps of the analysis were performed in accordance with the man-

manufacturer's protocols for both tests with additional primer targets the human beta-globin gene to provide control for cell adequacy, extraction and amplification.

Both of the tests have CE mark approval for use as the diagnostic tests in Europe.

RESULTS:

In the population studied, cell abnormalities were cytological diagnosed in 77.05% of HPV positive samples, but cellular abnormalities were also found in 25.8% of the samples that tested HPV negative.

Genotyping by Linnear Array test showed all together 15 different genotypes of HPV found in this study. Infection with HPV type 16 proved to be the most common (30.6%), followed in descending order of frequency by HPV- 51, -52, -31, -18, -58, -56, -45 and type 73 in 20.1%, 16.8%, 15.9%, 9.7%, 7.9%, 7.8%, 5.3% and 2.6% respectively. Mono-type infection (infection with single type of virus) was proven with Linnear Array test in 30.4% (82/269) cases, but in most patients - 69.5% (187/269), infection was caused by multi-type infection.

"Figure 1. about here?."

From each patient, samples were used for Amplicor test as well as for Linnear Array test. Between the tests used, difference was noticed in 44 of all samples -(12.9%; 44/339), as is shown in table 1. It is visible, that in spite of negative Amplicor result, in the 32 samples (72.2%; 32/44) certain types of HPV were found by Linnear Array Test. Seventeen of them were in high risk group, and 15 of them were of low risk types.

In twelve samples no viral type was found in spite of a positive Amplicor Test.

"Table 1. about here?."

Among Linnear positive and Amplicor negative patients, ten patients had cytological abnormalities (low grade lesion as a cut-off value for abnormality): eight of them had low grade epithelial abnormalities (LSIL) and two had high grade lesions (HSIL-CIN2 and HSIL-CIN3) proven by histology.

"Table 2. about here?."

Among Amplicor positive samples, 27.2% (12/44) of them were false positive, since in 12 samples none of the HPV types was detected by Linnear Array Test.

Adding the cytological data to the results of the patients with the proven HPV infection by Amplicor Test revealed 27.2% (53/195) of HPV positive patients in group with no cell abnormalities. In the group of patients with cellular abnormalities, HPV infection was confirmed among ASCUS, LSIL, HSIL and carcinoma cases in the samples, in 82.9% (39/47); 88% (103/117), 94.5% (52/55) and 100%, respectively. Results using Linnear Array test are slightly different, as is shown in table 2.

"Table 3. about here?."

The sensitivity as well as specificity of these tests has been observed to be somewhat dissimilar, though not statistically significantly ($p > 0,05$).

"Picture 2. About here?."

DISCUSSION:

Ever since G. Papanicolaou described a method for detecting cancer cells in the cervical smears in the first half of the 20th century, this method called the Pap test, has been established as a gold standard for early detection of the precancerous and cancer lesions and soon has become a method of choice for cervical cancer screening programs all over the world⁵. There are lot of provide evidence that cervical lesions are associated with persistent infection with oncogenic types of HPV. Screening programs provide a way to identify the potential for disease in apparently asymptomatic patients, and since infection with HPV is completely asymptomatic, screening for the as-

sociated disease is of an extreme importance.

Currently, the recommendations for HPV DNA testing for cervical cancer screening include triaging patients with equivocal or low-grade cytological abnormalities⁷, co-testing in combination with cytology in patients over 30 years of age to guide referral for colposcopy, and it's recommended for prediction of the therapeutic outcome after treatment of cervical intraepithelial neoplasia (CIN) lesions^{8,9}.

Two HSIL, eight LSIL and four ASCUS lesions were found in the present analysis, among patients with discrepancy between the used tests. Similar results were found by other authors, as the Linnear Test has been used as a control test in cases of discrepancies between commercial HC2 (Hybrid Capture 2) test and Amplicor test^{10,11}. The difference between the tests may be explained by the difference in the number of viral copy that has been used by tests as well as determined positive diagnostic rate¹².

In this study, HPV infection was found in 27% of the patients with no cytology abnormalities. Prevalence of high risk HPV types was reported by many authors and is up to 10% of women in the general screening population with no cervical abnormalities, but results showed a lot of diversities. Correct result in detection of the HPV infection varies and depends on several variables like: exact patients population involved in the study¹³, cytological status of the selected population, their age, occupation, and among others things -on the assay used for the HPV discovery^{10,14-18}.

After epidemiologic classification of HPV types associated with cervical cancer, made by Munoz et al.¹⁸, many of performed meta-analyses confirmed these results all over the world, and revealed five most common genotypes as follows: type 16, 18, 45, 31 and 33^{14,17-20}.

Analyses also exposed slight differences in the geographical distribution of hr-HPV genotypes around the globe, and the existence other oncogenic types of HPV which are responsible for cervical diseases^{19,20}. Despite of those differences, there are far more similarities. Worldwide distribution of HPV types 16 and 18 was found in almost 70% of the cervical cancers, as well as in precancer lesions (HSIL)²¹.

Recent studies by Snijders et al. have shown that HPV testing is about 45% more sensitive than cytology alone in detecting high-grade lesions, so it has been concluded that this is a good enough reason to use HPV testing alone for primary cervical screening²².

Data from the ATENA study, introduced and clinically validated the Cobas HPV Test (Roche Molecular System. Inc) as a potential test for primary screening in women over 21 years. This study showed that risk of HSIL-CIN 2 or higher lesion in the population of HPV-type 16 positive women with no cervical abnormalities is up to 13.6%, and that among hr HPV negative women, estimated risk of developing HSIL is 0.3% and 0.8% for CIN 2 and CIN 3, respectively²³⁻²⁵.

The ATENA study is based on an earlier US study by Khan et al. where the authors provide an option for managing patients ≥ 30 years with negative cytology and positive HPV type 16/18²⁶. These patients should be referred to colposcopy, while those women positive for one of the other high risk types should be retesting within 12 months with cytology and another HPV testing.

In the Europe, Netherlands will probably be the first European country to adopt HPV testing as the primary tool for organised cervical screening. The Dutch Council for Health Care has advised the Ministry of Health Care to introduce primary HPV testing in the cervical cancer screening program while cytology remains the option of choice for triage and management of high risk HPV positive patients²⁷.

In conclusion, it can be generally recommended that HPV genotyping should be included in the cervical cancer screening program in every country because the precise data on hr-HPV genotype distributions have implications for follow-up protocols in screening programs but also in achieving the highest sensitivity and specificity possible for diagnostic tests used for screening and for assessing the impact of a vaccine program.

Consensus on the number and types of HPV should exist in every country and has not been reached yet, hence further and extensive investigation is recommended.

TABLES AND PICTURES:

FIGURE 1. Genotype distribution and type of HPV infection in the study

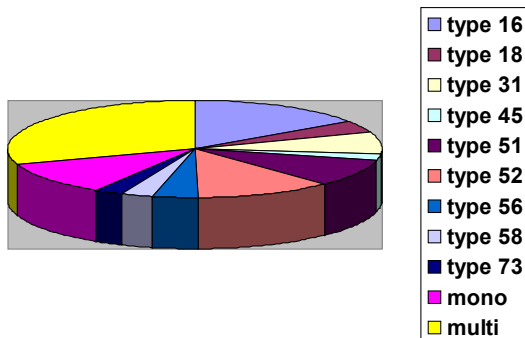


TABLE 1. Difference between Amplicor and Linnear Array Tests

Amplicor test	Linnear Array Test		Total positive	Total negative	TOTAL
	High-risk type	Low-risk type			
Positive	145	23	168	12	180
Negative	17	15	32	57	89
TOTAL			200	69	269

Se=84%; Sp=82,6%; PPV= 93,3%; NPV= 64% for Amplicor test

**Se= 93,3%; Sp=64%; PPV= 84%; NPV= 82,6% for Linnear Array test

(Se =sensitivity;Sp=specificity;PPV=positive predictive value; NPV= negative predictive value)

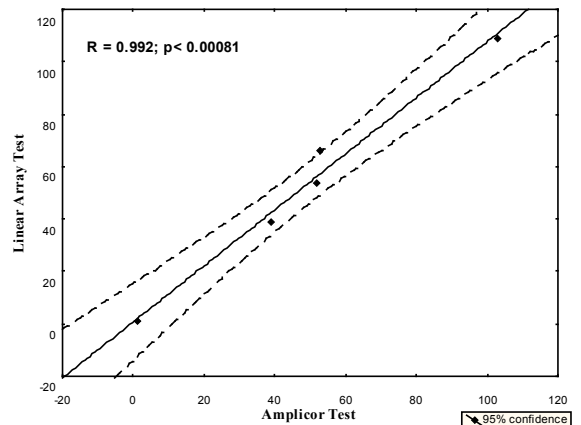
TABLE 2. Relationship between cytology data and HPV results with Amplicor Test and Linnear Array Test

Cytology result	Amplicor test		Linnear Array test		TOTAL
	Positive	Negative	Positive	Negative	
No abnormalities	53	66	66	53	119
TOTAL No abnormalities	53	66	66	53	119
ASCUS	39	8	39	8	47
LSIL	103	14	109	8	117
HSIL (cin2+3)	52	3	54	1	55
Ca	1	--	1	--	1
TOTAL with abnormalities	195	25	203	17	220

TABLE 3. Results of Odds ratio test and Chi-square test for cytology data between Amplicor Test and Linear Array Test

Cytology result	Odds ratio test		Chi-square test		
	Odds ratio	95 % CI	95 % CI	c ²	P
No abnormalities	0.64	0.39 - 1.08	-7.63 to 23.03	0.599	0.4391
ASCUS	1.00	0.34 - 2.93	-	-	-
LSIL	0.54	0.22 - 1.34	-4.91 to 11.51	0.318	0.5728
HSIL (cin2+3)	0.32	0.03- 3.19	-3.59 to 10.79	0.220	0.6391
Ca	-	-	-	-	-
TOTAL (with abnormalities)	0.65	0.34 - 1.25	-	-	-

FIGURE 2. Results of linear correlation obtained for cytology data between Amplicor Test and Linear Array Test



REFERENCES:

- [1] Nobbenhuis MA, Helmerhorst TJ, van den Brule AJ, Rozendaal L, Voorhorst FJ, Bezemer PD, Verheijen RH, Meijer CJ (2001). "Cytological regression and clearance of high-risk human papillomavirus in women with an abnormal cervical smear", *ELSEVIER, Lancet*, 358: 1782-3.
- [2] Zielinski GD, Snijders PJ, Rozendaal L, Voorhorst FJ, van der Linden HC, Ronsink AP, de Schipper FA, Meijer CJ (2001). "HPV presence precedes abnormal cytology in women developing cervical cancer and signals false negative smears", *NATURE PUBLISHING GROUP, Br J Cancer*, 85: 398-404.
- [3] Kjaer SK, van den Brule AJ, Paull G, Svare EI, Sherman ME, Thomsen BL. (2002) "Type specific persistence of high risk human papillomavirus (HPV) as indicator of high grade cervical squamous intraepithelial lesions in young women: population based prospective follow-up study". *BMJ*, 325: 572.
- [4] Bosh FX, Manos MM, Muñoz N, Meijer CJ, Shah KV (2002) "The causal relation between papillomavirus and cervical cancer". *OXFORD UNIVERSITY PRESS, J Clin Pathol*, 55: 244-65
- [5] Doorbar J. "HPV infection, the nature of the infected cell and the outcome of cervical disease"(2012). *HPV today*, 25:5.
- [6] Krivak Bolanča I, Vraneš J.(2010) "Diagnostic methods and techniques in preventing cervical carcinoma Part I: Conventional cytology and new cytological methods". *ZENICA : MEDICAL ASSOCIATION OF ZENICA-DOBOJ CANTON*, *Med Glas*, 7(1): 12-17.
- [7] Wright TC jr, Massad LS, Dunton CJ, Spitzer M, Wilkinson EJ, Solomon D. (2007). "Consensus guidelines for the management of women with abnormal cervical cancer screening tests". *ELSEVIER, Am J Obstet Gynecol*, 197(4): 346-55.
- [8] Quigley NB, Potter NP, Chivukula M, Knight MZ, Welch JR, Olson MC.(2011) "Rate of detection of high-risk HPV with two assays in women ≥ 30 years of age".*ELSEVIER, J Clin Virology*, 52(1): 23-27.
- [9] ACOG. "Practice Bulletin No.109 cervical cytology screening" (2009). *WOLTERS KLUWER HEALTH, Obstet Gynecol*, 114(6):1409-20.
- [10] Koidl C, Bozic M, Hadzisejdic I, Grahovac M, Grahovac B, Kranewitter W, Marth E, Kessler HH. (2008) "Comparison of molecular assays for detection and typing of human papillomavirus". *ELSEVIER, Am J Obstet Gynecol*, 199: 144.
- [11] Hardie A, Moore C, Patnick J, Cuschieri K, Graham C, Beadling C, Ellis K, Frew V, Cubie HA.(2009) "High-risk HPV detection in specimens collected in SurePath preservative fluid: comparison of ambient and refrigerated storage". *WILEY, Cytopathology*, 20: 235-41.
- [12] Hawkins DM, Garrett JA, Stephenson B.(2001) "Some issues in resolution of diagnostic tests using an imperfect gold standard". *WILEY, Stat Med*, 20: 1987-2001.
- [13] Evander M, Edlund K, Gustafsson A, Jonsson M, Karlsson R, Rylander E, Wadell G.(1995)

- "Human papillomavirus infection is transient in young women: a population-based cohort study". *ELSEVIER, J Infect Dis*, 171: 1026-30.
- [14] Jacobs MV, Walboomers JM, Snijders PJ, Voorhost FJ, Verheijen RH, Meijer CJ.(2000) "Distribution of 32 mucosotropic HPV types in women with cytological normal cervical smears: the age-related patterns for high-risk and low-risk types". *WILEY. Int J Cancer*, 87: 221-7.
- [15] Husnjak K, Grce M, Magdić L, Pavelić K.(2000) "Comparison of five different polymerase chain reaction methods for detection of human papillomavirus in cervical cell specimens". *ELSEVIER, J Virol Methods*, 88(2): 125-34.
- [16] Milutin Gasperov N, Sabol I, Matovina M, Spaventi S, Grce M.(2008) "Detection and typing of human papillomaviruses combining different methods: polymerase chain reaction, restriction fragment length polymorphism, line probe assay and sequencing". *SPRINGER, Pathol Oncol Res*, 14(4): 355-63.
- [17] Bao YP, Li N, Smith JS, Qiao YL.(2008) "Human papillomavirus type distribution in women from Asia: a meta-analysis". *WOLTERS KLUWER HEALTH, Int J Gynecol Cancer*, 18: 71-9.
- [18] Munoz N, Bosch FX, de Sanjose S.(2003) "Epidemiologic classification of human papillomavirus types associated with cervical cancer". *MASSACHUSETTS MEDICAL SOCIETY, N Engl J Med*, 348: 518-27.
- [19] de Sanjose S, Diaz M, Castellsagué X, Clifford G, Bruni L, Muñoz N, Bosch FX.(2007) "Worldwide prevalence and genotype distribution of cervical HPV DNA in 157,879 women with normal cytology: a meta-analysis of 78 studies". *ELSEVIER, Lancet Infect Dis*, 7: 453-59.
- [20] Clifford GM, Smith JS, Aguada T.(2003) "Comparison of HPV types distribution in high-grade cervical lesions and cervical cancer: a meta-analysis". *NATURE PUBLISHING GROUP, Br J Cancer*, 89: 101-6.
- [21] de Sanjose S, Quint WG, Alemany L, Geraets DT, Klaustermeier JE.(2010) "Human papillomavirus genotype attribution in invasive cervical cancer: a retrospective cross-sectional worldwide study". *ELSEVIER, Lancet Oncol*, 11: 1048-56.
- [22] Snijders PJF.(2011) HPV today, Newsletter on Human Papillomavirus, 24:1-3.
- [23] Castle PE, Stoler MH, Wright TC Jr, Sharma A, Wright TL, Behrens CM.(2011) "Performance of carcinogenic human papillomavirus (HPV) testing and HPV16 and HPV18 genotyping for cervical cancer screening of women aged 25 years and older: a subanalysis of the ATHENA study". *ENSELVIER, Lancet Oncol*, 12(9): 880-90.
- [24] Wright TC Jr, Stoler MH, Sharma A, Zhang G, Behrens C, Wright TL.(2011) "ATHENA (Addressing THE Need for Advanced HPV Diagnostics) Study Group. Evaluation of HPV-16 and HPV-18 genotyping for the triage of women with high-risk HPV+ cytology-negative results". *OXFORD UNIVERSITY PRESS, Am J Clin Pathol*, 136(4): 578-86.
- [25] Stoler MH, Wright TC Jr, Sharma A, Apple R, Gutekunst K, Wright TL.(2011), "ATHENA HPV Study Group. High-Risk Human Papillomavirus Testing in Women With ASC-US Cytology. Results From the ATHENA HPV Study". *OXFORD UNIVERSITY PRESS, Am J Clin Pathol*, 135: 468-75.
- [26] Khan MJ, Castle PE, Lorincz AT, Wacholder S, Sherman M, Scott DR, Rush BB, Glass AG, Schiffman M.(2005) "The elevated 10-year risk of cervical precancer and cancer in women with human papillomavirus (HPV) type 16 or 18 and the possible utility of type-specific HPV testing in clinical practice". *OXFORD UNIVERSITY PRESS, J Natl Cancer Inst*, 97:1072-9.
- [27] Gök M, van Kemenade FJ, Heideman DA, Berkhof J, Rozendaal L, Spruyt JW, Beliën JA, Babovic M, Snijders PJ, Meijer CJ.(2012) "Experience with high-risk human papillomavirus testing on vaginal brush-based self-samples of non-attendees of the cervical screening program". *WILEY, Int J Cancer*, 130(5):1128-35.